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**TERT** promoter status and gene copy number gains: effect on TERT expression and association with prognosis in breast cancer

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**ABSTRACT**

Upregulation of the telomerase reverse transcriptase (**TERT**) gene in human cancers leads to telomerase activation, which contributes to the growth advantage and survival of tumor cells. Molecular mechanisms of **TERT** upregulation are complex, tumor-specific and can be clinically relevant. To investigate these mechanisms in breast cancer, we sequenced the **TERT** promoter, evaluated **TERT** copy number changes and assessed the expression of the **MYC** oncogene, a known transcriptional **TERT** regulator, in two breast cancer cohorts comprising a total of 122 patients. No activating **TERT** promoter mutations were found, suggesting that this mutational mechanism is not likely to be involved in **TERT** upregulation in breast cancer. The T349C promoter polymorphism found in up to 50% of cases was not correlated with **TERT** expression, but T349C carriers had significantly shorter disease-free survival. **TERT** gains (15-25% of cases) were strongly correlated with increased **TERT** mRNA expression and worse patient prognosis in terms of disease-free and overall survival. Particularly aggressive breast cancers were characterized by an association of **TERT** gains with **MYC** overexpression. These results evidence a significant effect of gene copy number gain on the level of **TERT** expression and provide a new insight into the clinical significance of **TERT** and **MYC** upregulation in breast cancer.

**INTRODUCTION**

The telomerase reverse transcriptase (**TERT**) gene encodes the catalytic subunit of telomerase that maintains telomere length. Telomerase activity occurs in more than 90% of cancers [1]. One of the hallmark features of tumor cells, it contributes to their growth advantage and survival, and upregulation of the **TERT** gene is the major mechanism of telomerase activation in human cancer [2]. In breast tumors, **TERT** overexpression was associated...
Several molecular events can modify TERT expression in cancer cells. Somatic mutations and functional single nucleotide polymorphisms (SNP) in the TERT promoter can lead to changes in the expression of the gene [6]. Overexpression of the oncogene MYC can directly enhance TERT expression by increased binding to the TERT promoter [7]. Increased copy number or amplification of the TERT gene can also result in TERT upregulation [8–10].

The TERT promoter is the most important element of telomerase expression because it harbors binding sites for numerous cellular transcriptional activators and repressors that directly or indirectly regulate gene expression. Recurrent somatic mutations in the TERT promoter were found in cancers of the central nervous system (43%), bladder (59%), thyroid (follicular cell-derived, 10%) and skin (melanoma, 29%) [11, 12]. In particular, two hotspot C228T and C250T mutations create binding motifs for transcription factor ETS2 and increase TERT transcriptional activity [13]. In addition, the T349C SNP, chr5:1,295,349 T>C (rs2853669) affects telomerase activity and telomere length [14, 15]. This polymorphism can be studied in both blood and tumor samples since the results are concordant [14, 16].

In breast cancer, only limited data are available concerning the incidence of mutations and SNP in the TERT promoter, TERT gains and their relationship with TERT upregulation and patient prognosis. To address these issues, we investigated the mutational status and genotype of the TERT promoter, TERT gene copy number and TERT and MYC mRNA expression in two breast cancer cohorts. The first cohort comprised 77 patients with triple-negative breast cancer (TNBC) from two recent neoadjuvant trials assessing the efficacy and toxicity of anti-EGFR antibodies combined with chemotherapy [17, 18]. The patients had been followed for 5 years. The second cohort consisted of a retrospective series of 45 patients treated with standard neoadjuvant chemotherapy (NCT). Most of them had common hormone-positive tumors and had been followed for more than 10 years.

RESULTS

TERT promoter status

We detected no activating TERT promoter mutations, including hotspot mutations C228T and C250T, in 77 pre-NCT breast tumor biopsies (cohort #1) and 45 post-NCT residual tumors (cohort #2). The T349C (rs2853669) SNP was identified in 42.3% of tumor biopsies and 48.9% of residual tumors (Table 1). In both #1 and #2 cohorts, most tumors with the variant C allele had a heterozygous 349 T/C genotype (37.2% and 44.4%, respectively) whereas the frequency of 349 C/C homozygotes was low (5.1% and 6.7%). Overall, the frequencies of T349C in the two cohorts were similar to those in the 1000 Genome database for Europe.

TERT copy number aberrations

We found that TERT gains were relatively frequent events in both cohorts. Twenty tumors (25.6%) in cohort #1 and seven tumors (15.6%) in cohort #2 had TERT gains. TERT losses were rare, with the alteration occurring in three tumors (3.9%) in cohort #1 and two tumors (4.4%) in cohort #2.

 Associations of the T349C genotype and TERT gene copy number gains with TERT and MYC expression

We investigated whether T349C genotype and TERT gains were associated with changes in the expression of the TERT gene (Figure 1A–1B). The results were similar in the two cohorts. Tumors with 349 T/C or C/C polymorphism had increased expression of TERT, but the difference from the 349 T/T cases was not significant. In contrast, TERT gene copy number gains were associated
with a highly significant increase in gene expression as compared to cases without TERT gain. Of note, TERT gains were more frequent in the presence of the T349C SNP. In cases with 349 T/C or C/C genotypes, the relative risk of a TERT gain increased 3.2-fold ($P = 0.0036$, 95% confidence interval from 1.5 to 6.9) in cohort #1 and 5.7-fold in cohort #2 ($P = 0.11$, 95% confidence interval 1.0-32.0), as compared to the risk in 349 T/T cases.

In parallel, we investigated if the presence of TERT T349C polymorphism or gain was associated with the expression levels of MYC oncogene, a strong positive regulator of TERT expression (Figure 1C–1D). We found no association between the presence of the T349C SNP and MYC expression. In contrast, a significantly higher expression of the MYC gene were observed in cases with TERT gains (cohort #1, $P < 10^{-3}$; cohort #2, $P = 0.015$). Overall, the levels of TERT and MYC expression were significantly correlated in both cohorts (Pearson $r = 0.696$, $P < 10^{-7}$ in cohort #1 and $r = 0.542$, $P < 10^{-3}$ in cohort #2).

Clinical significance of the T349C SNP and TERT gene copy number gains

The presence of the T349C SNP (349 T/C and C/C genotypes) in cohort #1 was not significantly associated with clinicopathologic characteristics such as tumor size,
tumor grade, nodal status and pathologic response to NCT (Figure 2A). However, the rate of incomplete pathologic responses tended to be higher in T349C cases than in patients with T349T (69% vs. 51%, \( P = 0.12 \)). Patients with the T349C genotype had significantly shorter DFS and a trend toward a shorter OS (Figure 2B–2C).

In cohort #2, the T349C SNP was not associated with clinicopathologic characteristics (Figure 3A). T3-4 tumors were more frequent in T349C patients than in those with 349 T/T genotype (43% vs 23%), but not to a level of significance (\( P = 0.14 \)). Survival analysis showed, as in cohort #1, a significant effect of T349C on DFS but not on OS (Figure 3B).

Patients with \( TERT \) gains in cohort #1 had a trend toward a higher proportion of SBR grade III tumors (\( P = 0.13 \)) and a significantly higher rate of incomplete pathologic responses (\( P < 0.01 \), Figure 4A). No association with other clinicopathologic characteristics was observed.

Both DFS and OS were significantly shorter in patients with \( TERT \) gains (Figure 4B–4C).

In cohort #2, the number of cases with \( TERT \) gains was small (n=7) and the distribution of clinicopathological features among cases with and without gains was not significantly different (Figure 5A). However, the occurrence of a \( TERT \) gain was significantly related to shorter DFS and OS (Figure 5B–5C).

**MYC expression levels impact the prognostic value of \( TERT \) gains**

Univariate Cox analysis for DFS in cohort #1 showed that the presence of the T349C genotype and \( TERT \) gain increased significantly the risk of tumor recurrence with HR of 2.8 and 3.8, respectively (Table 2). High \( MYC \) expression (> median) also had a significant and negative effect on DFS (HR of 3.5). Importantly, very high levels

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**Figure 2:** Distribution of clinicopathologic characteristics: tumor size, tumor grade, nodal status and pathologic response (A) disease-free (B) and overall survival (C) in cohort #1 according to T349C status. Patients with 349 T/C or C/C alleles had significantly shorter DFS and a trend toward a shorter OS.
of MYC expression (upper quartile, > 421) were associated with a much higher risk of tumor recurrence (HR of 9.4, 95%-CI: 3.2-27.2, P < 0.0001). As expected, incomplete pathologic responses to NCT according to Chevallier’s classification had a significant negative prognostic effect (HR of 3.6). When adjusted for pathologic responses, TERT gain and MYC overexpression remained significant prognostic factors (Table 2). As mentioned above, tumors with TERT gain frequently had MYC overexpression, and it was assumed that the presence of both abnormalities would confer a worse prognosis. Indeed, the combination of MYC overexpression with TERT gains was associated with the shortest DFS and OS, whereas lower MYC levels annulled any negative effect of TERT gain on DFS and OS (Figure 6A–6B). In cases without TERT gain, MYC levels were not prognostic (Figure 6C–6D).

**DISCUSSION**

Telomerase activation protects malignant cells from apoptosis and senescence by preventing further telomere loss [2, 3, 19]. Regulation of telomerase activation in cancer is complex and multifactorial and involves transcriptional activation of the TERT gene [10].

Activating somatic mutations of the TERT promoter have been reported in various cancers [11, 12]. The two most common mutations, C228T and C250T, map -124 and -146 bp upstream of the TERT ATG site (chr5: 1,295,228 C>T and 1,295,250 C>T, respectively) and create binding sites for ETS/TCF transcription factors, which leads to a two- to four-fold increase in transcriptional activity [13, 20]. These mutations were not found in limited breast cancer series [11, 12]. Our results from a total of 122

![Figure 3: Distribution of clinicopathologic characteristics: tumor size, tumor grade, nodal status and pathologic response (A) disease-free (B) and overall survival (C) in cohort #2 according to T349C status. Patients with 349 T/C or C/C alleles had significantly shorter DFS but no difference in OS.](image-url)
breast tumors evidenced no TERT promoter mutations, suggesting that this mutational mechanism is not likely to be involved in TERT upregulation in breast cancer.

The variant C-allele of the rs2853669 (T349C) polymorphism disrupts an ETS2-binding site (EtsA) in the proximal region of the TERT promoter [14]. Although decreased binding of transcription factor ETS2 can modify the expression of the gene, opinions differ on its effect on the level of TERT expression in cancer cells. Our results from cases of breast cancer did not show any significant effect of the T349C SNP on TERT expression. Previous studies showed an association of the T349C genotype with reduced TERT expression in non-small cell lung cancer, bladder cancer and glioblastoma [16, 21, 22]. However, a recent report on four hepatocellular carcinoma cell lines described increased luciferase TERT promoter activity in the presence of the T349C SNP [23]. The T349C variant site is close to (2 bp downstream) the binding site of transcriptional repressor E2F1, and T/C and C/C variants seem to inhibit E2F1 binding to the TERT promoter. In the presence of activating TERT mutations, this SNP was suggested to enhance TERT transcription levels by blocking E2F1 binding to its promoter. These recent findings suggest that the cooperation of native and changed (mutations, SNP) binding sites for multiple transcriptional factors in the TERT promoter could regulate TERT expression and result in cancer-specific effects of SNP.

There has been much debate about the clinical effect of the T349C SNP on the survival of patients with malignancies. Several studies reported its positive prognostic value in the context of activating TERT

**Figure 4:** Distribution of clinicopathologic characteristics: tumor size, tumor grade, nodal status and pathologic response (A) disease-free (B) and overall survival (C) in patients from cohort #1 with and without TERT gene copy number gain. Patients with TERT gains had a significantly higher rate of incomplete pathologic responses and significantly shorter DFS and OS.
promoter mutations in bladder cancer and clear cell renal cell carcinoma [22, 24, 25]. However, more recent reports have shown that this SNP has a negative effect in glioblastoma and liver cancer patients bearing activating TERT promoter mutations [23, 26]. Among acute myeloid leukemia patients without TERT promoter mutations, T349C carriers had significantly shorter survival times [27]. Our study shows for the first time a negative effect of the T349C SNP on survival in breast cancer patients in the absence of activating somatic mutations. However, this SNP was significantly associated with TERT copy number gains in our patient series.

Increased TERT gene dosage was seen in about 30% of various human tumor-derived cell lines, including breast cancer cells [28]. We found TERT copy number gains in more than 20% of primary breast tumors examined. Previous studies yielded conflicting results regarding the effect of TERT gain on TERT mRNA expression. TERT gene gains were associated with higher TERT mRNA expression in non-small-cell lung adenocarcinoma and Merkell cell carcinoma [29, 30] and with high TERT protein expression in breast cancer and cervical carcinomas [28, 31]. However, no significant association was found in colorectal carcinomas and squamous-cell lung cancer [30, 32]. Our observation of a highly positive correlation between the TERT gene copy number and its mRNA level suggests that TERT gain has a functional effect on TERT transcription in breast cancer.

We showed that TERT gains identified in pre-NCT biopsies were predictive of incomplete pathological responses and shorter survival. In addition, TERT gain was a significant predictor of worse survival when assessed in

![Figure 5](image)

*Figure 5*: Distribution of clinicopathologic characteristics: tumor size, tumor grade, nodal status and pathologic response (A) disease-free (B) and overall survival (C) in patients from cohort #2 with and without TERT gene copy number gain. Patients with TERT gains had a significantly higher rate of incomplete pathologic responses and significantly shorter DFS and OS.
Table 2: Cox analysis for disease-free survival in breast cancer cohort #1

<table>
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<th>Parameters</th>
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<th>Adjusted*</th>
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<tr>
<td></td>
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<td>P</td>
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<td>TERT gain (yes vs no)</td>
<td>3.8 (1.4-10.3)</td>
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<tr>
<td>TERT T349C (yes vs no)</td>
<td>2.8 (1.0-7.7)</td>
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<td>MYC (high vs low)**</td>
<td>3.5 (1.1-10.8)</td>
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<td>Chevallier’s classification (3-4 vs 1-2)</td>
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<td>0.048</td>
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*Adjusted for treatment response according to Chevallier’s classification: incomplete pathologic response, classes 3-4, vs complete pathologic response, classes 1-2.
**High > median, low ≤ median (181)
Hazard ratio, HR; 95% confidence interval, 95% CI.

Figure 6: The combination of TERT gene copy number gains and MYC overexpression was associated with the shortest DFS and OS in cohort #1. Lower MYC levels annulled the negative effect of TERT gains on DFS and OS (A, B). In cases without TERT gains, MYC levels had no effect on patient survival (C, D).
residual post-NCT tumors. To our knowledge, only two studies have reported a negative effect of TERT gains on survival in non-small cell lung carcinomas and melanomas [30, 33]. For the first time, we show that TERT gain is also a negative prognostic marker associated with higher risk of disease recurrence or death from breast cancer and could potentially be used to adapt treatment strategies.

TERT upregulation correlates with increased telomerase activity. Inhibition of telomerase in breast tumors might potentiate the effects of standard anti-cancer treatments [34]. This opens up the perspective of using telomerase inhibitors in patients resistant to NCT. Preclinical studies with the telomerase inhibitor GRN163L showed that it can restore sensitivity of HER2+ breast cancer cell lines to trastuzumab and so patients with HER2+ breast cancers were recruited into a phase I clinical trial (NCT 01265927). In combination with chemotherapy, GRN163L was used in a phase II clinical trial in patients with locally recurrent or metastatic breast cancer (NCT01256762). These clinical trials have been completed but the study results are not yet available [34].

Telomerase reactivation and/or TERT upregulation correlate with MYC overexpression [35, 36]. In line with these findings, we observed that MYC and TERT expression levels are significantly correlated. MYC is a primary transcription factor of the TERT gene and stimulates its promoter activity [37, 38]. In breast cancer cells, MYC binds to an ETS2 site (EtsA) in the TERT gene promoter and increases TERT expression levels [15]. As mentioned above, this binding site is disrupted by T349C variants, but we observed no significant changes in TERT or MYC expression according to the presence or absence of this SNP.

In contrast, MYC overexpression was strongly associated with the presence of TERT gains. Interestingly, a recent study showed that TERT regulates MYC ubiquitination, stabilization and binding to its target gene promoters [39]. This feed-forward transcriptional loop between MYC and TERT is functionally critical for oncogenesis. In our study, we found that MYC overexpression often coincided with TERT gains. Enhanced MYC stability and function in cells with high TERT and MYC levels could potentiate MYC-dependent oncogenesis.

By combining the data on TERT gain and MYC overexpression we were able to isolate a particularly bad prognosis subgroup of breast cancer patients. MYC is considered to promote tumor cell survival, proliferation and progression in breast cancer [40, 41] and regulates the expression of 13 different poor-outcome cancer signatures [42]. We found that MYC overexpression had a negative prognostic value in the whole patient population and a particularly significant effect within the subgroup of patients with TERT gains. A concomitant upregulation of TERT and MYC identified patients with a high risk of breast cancer recurrence. In contrast, lower MYC expression levels were associated with a favorable prognosis, despite the presence of TERT gains.

In conclusion, we found no activating TERT promoter mutations in 122 breast cancer patients. The T349C TERT promoter SNP was not significantly associated with TERT expression but T349C carriers had shorter survival. Notably, TERT gene copy number gain was significantly related to TERT upregulation and, in association with MYC overexpression, characterized particularly aggressive disease. These results show a significant effect of gene copy number gain on TERT expression level and provide a new insight into the clinical significance of TERT and MYC upregulation in breast cancer.

**MATERIALS AND METHODS**

**Patients and samples**

Cohort #1 comprised 77 patients with operable, stage II–III TNBC enrolled in two neoadjuvant trials evaluating the efficacy and toxicity of anti-EGFR antibodies plus chemotherapy: panitumumab combined with a standard neoadjuvant anthracycline/taxane-based [17] and cetuximab combined with docetaxel [18]. After NCT, pathologic complete responses (classes 1 and 2 according to Chevallier’s classification) were observed in 32 of 77 cases (41.6%). The remaining 45 (58.4%) patients had incomplete pathologic responses (class 3, N = 39 and class 4, N = 6). The median follow-up in this cohort was 3.8 years (range, 0.9-5.5 years). During follow-up, 16 (20.8%) patients relapsed and 11 (14.3%) died from breast cancer. Cohort #2 consisted of archived samples of residual post-NCT tumors from 45 women diagnosed with invasive breast cancer and treated with NCT protocols [43] between 1996 and 2010 at the Jean Perrin Cancer Center (Clermont-Ferrand, France). Following NCT, patients underwent appropriate surgery and radiotherapy. Those with important residual disease received adjuvant chemotherapy. Menopausal patients with hormone receptor-positive tumors received tamoxifen for 5 years. The mean follow-up was 9.1 years (range, 1.1-15.9 years). During this period, 20 (44.4%) patients relapsed and 14 (31.1%) died from breast cancer. In cohort #1, the study was performed on pre-treatment tumor biopsies and in cohort #2 on post-NCT residual tumor samples cryopreserved in the Biological Resources Center BB-0033-00075. DNA and RNA were simultaneously extracted with the AllPrep DNA/RNA/miRNA Universal Kit (Qiagen, Courtaboeuf, France) according to the manufacturer’s instructions. The study was approved by the institutional review board.

**Gene expression analysis**

Total RNA was converted to cDNA by reverse transcription with Superscript II reverse transcriptase (Invitrogen, Cergy-pontoise, France) according to the
manufacturer’s instructions. The expression of TERT was quantified by real-time RT-PCR in the Lightcycler 480 System (Roche Diagnostics, Meylan, France) as described previously [44, 45]. MYC was quantified with primers described in the literature [46]. The normalized copy numbers were expressed as the ratio between transcript copy numbers of the target and control (B2M) genes multiplied by 100.

**TERT promoter sequencing**

The amplification of genomic DNA for the TERT promoter region (267 pb) was performed with the following primers: forward 5’-CCGGGCTCCCAGTGGATT-3’ and reverse 5’-TTCCCACGTGCAGCAGCAG-3’. PCR conditions consisted of an initial heating at 95°C for 15 minutes followed by 40 cycles at 95°C for 30 seconds, 60°C for 20 seconds and 72°C for 30 seconds. Amplified PCR products were sequenced with BigDye v1.1 on the ABI 3500 Genetic Analyzer (Applied Biosystems, Courtaboeuf, France).

**TERT copy number quantification**

TERT gene copy number was quantified by Quantitative Multiplex Fluorescent Polymerase Chain Reaction (QMF-PCR) analysis. QMF-PCR was performed on a TProfessional thermocycler (Biometra, Archamps, France) as previously described [47, 48]. Primers were designed for three fragments of the TERT gene on chromosome 5p (5’UTR, Exon 9 and 3’UTR) to cover the entire gene. The 5’UTR region is very GC-rich and was difficult to amplify. We therefore had to limit the analysis to exon 9 and 3’UTR. Seven genes on chromosomes 2p, 4p, 7q, 10q, 11p, 11q (PVRL1, BOD1L, RET, ZNF638, AGBL2, CFTR and POR) were co-amplified as controls. Primers and technical details are described in Supplementary Methods. The fluorescence intensities of PCR products were correlated with the copy number of the relevant exons. Fourteen control DNAs were included in each experiment: 12 normal DNAs and 3 DNAs with known TERT copy number (loss, normal and gain, as determined by array-comparative genomic hybridization). A dosage quotient was calculated relative to all the other amplified exons in patients and controls. The range of ratios corresponding to two copies of TERT gene was set between 0.8 and 1.2 (mean +/- 3 standard deviations of values obtained in normal DNA samples). Ratios > 1.2 were considered as gains and < 0.8 as losses.

**Statistical analysis**

Standard tests (Kruskal-Wallis H test, ANOVA, Student’s t-test, chi-squared test, Pearson’s correlation) were used to study the relationship between characteristics. Disease-free survival (DFS) was measured from diagnosis until the first relapse (local or distant) and overall survival (OS) from the start of NCT until the last follow-up report. Survival curves were established by the Kaplan-Meier method and compared with the log-rank test.

**Author contributions**

MGB performed the experimental work, analysed the data and drafted the manuscript. LV, PC, EEP, MMD, AC and MP contributed to the experimental work. FK performed statistical analyses. CA, YJB, MAMR and PV contributed to the data analysis. AT and FPL designed and supervised the study. AT wrote the final manuscript. All authors contributed to the final version of the manuscript.

**CONFLICTS OF INTEREST**

The authors declare no conflicts of interest.

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